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Schiff bases of chitosan

An overview

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Abstract

Chitosan is a polysaccharide that has amino and hydroxyl groups and other functional groups that enable functionalization. Schiff bases, which can be identified using ¹H NMR, FTIR spectrometry and thermal analysis, can be created when the C2-NH2 group reacts with primary amines.

Keywords: chitosan, Schiff base, spectral methods, thermal analysis

1. INTRODUCTION

Chitosan is a biopolymer comprised of N-acetylated glucosamine and glucosamine units connected by β -(1–4) glycosidic bonds (Scheme 1). It is obtained by the deacetylation of chitin. Chitin can be found in the shells of crustaceans, insects, and various other organisms such as fungi, algae, and yeast [1].

Chitosan molecules contain several functional groups, including C₃–OH, C₆–OH, C₂–NH₂, and acetyl amino and glycoside bonds. The acetylamino and glycosidic bonds are equally stable and not easily

broken. The C₃–OH group is a secondary hydroxyl group that cannot rotate freely and has significant steric hindrance, making it difficult to react. On the other hand, the C₆–OH and C₂–NH₂ groups are more active chemically, and their presence in chitosan allows for the introduction of other functional groups through various molecular design strategies. They can participate in metal coordination, chemical coupling, chemical crosslinking, graft copolymerization or alkylation reactions. The C₆–OH group can be esterified and carboxymethylated, too. The C₂–NH₂ groups can be acylated and Schiff bases are formed. Glycosidic bonds can be cleavaged by degradation [2].

This review article centres on the examination of chitosan's Schiff bases, along with their characterization and diverse applications.

1. Schiff bases of chitosan characterization

Schiff base compounds which have an imine group (-RC=N-) can be produced by the reaction of a primary amine with an active carbonyl. Chitosan, a polymer containing amino groups in its chain, is susceptible to various chemical modifications, such as the creation of Schiff bases via the reaction with aldehydes and ketones (Scheme 1).



Scheme 1. Synthesis of Schiff bases of chitosan

Characterization techniques FTIR and ¹H NMR spectroscopy were used to characterize the biopolymeric Schiff bases of chitosan.

Chitosan and Schiff bases of chitosan show an intense band in the region of 3400 cm⁻¹ assigned to the stretching vibrations of the -OH and –NH₂ groups of their structure. A band between 3000 and 2880 cm⁻¹ was assigned to symmetrical and asymmetric C-H stretch. Other characteristic bands of chitosan are shown at 1690-1630 cm⁻¹, 1600-1560 cm⁻¹, 1430-1380 cm⁻¹ and 1200-1000 cm⁻¹ which were attributed to C=O stretching of the acetamide group (amide band I), N-H stretching of acetamide group (amide band II), C-N deformation of amino group (amide group III) and O-C-O stretching of the glycopyranoside ring, respectively. The spectra of chitosan-Schiff bases derivatives show a new band at 1650-1560 cm⁻¹ due C=N vibrations. The bands ranging from 1400 to 1500 cm⁻¹ are due to the C-C stretching vibrations of the aromatic ring. The absence of the characteristic band of aldehyde in the region 1660–1730 cm⁻¹ indicates that there is no traceable residue of free aldehydes [1-9].

A significant change in the spectra was observed when comparing the Schiff bases to chitosan, with distinct ¹H NMR signals characterizing each [5-6, 8-9]. The chitosan spectra and the Schiff base spectra exhibited a singlet between 1.23 and 1.87 ppm, attributed to the acetyl proton, and a singlet between 1.91 and 2.98 ppm, corresponding to the (H-2) proton of the pyranose ring. On the other hand, the Schiff bases demonstrated a sharp singlet between 8.78 and 9.79 ppm due to the imine group's proton (–N=CH–), and different signals between 6.0 and 10.2 ppm, mainly resulting from the aromatic rings present in the Schiff bases.

Chitosan-Schiff bases were characterized by thermogravimetric analysis [1-5, 6-8]. Thermogravimetry of Schiff bases derived from chitosan shows that they have nearly the same decomposition temperature as the chitosan, indicating that Schiff bases are thermally stable. The studied Schiff base of chitosan has water elimination up to 120°C. The second degradation step is observed around 200°C-300°C, due to the decomposition of the polymeric chains followed by the thermal degradation of a new material formed by the residual of the second stage of the degradation process.

f base of chitosan		Main IF	s abso	rption	n band	ls (cm	1))		NMR	peaks	(wdd/Q)	TGA	Ref
												-	analysis	
		VNH2 VOH VC-H	VC=0	NNH	VC-N	Vglyc	VC=N	VC=C	H- amide	H-2	H- imine	H- _{Ar}	Temp. (°C)	I.
se of robe	chitosan with 4- nzaldehyde	3200- 3200- 2919 3600 3600 2879	1650	1575	1380	1150				L	I.	ı	~100°C 220-350°C t>350°C	[1]
hitos	an with benzophenone	3200- 3200- 2919 3600 3600 2879	1650	1575	1380	1150	1	1		I	I.	I	~100°C 220-350°C t>350°C	[1]
tosaı	- S-andole-3-	3200- 3200- 2960 3400 3400 1933	1645	1565	1370	1000- 1200	ı	ı	ı	ı	I	I	ambient- ~100°C 140-200°C	[3]
tosai carbé	n with 1H-pyrazolo[3,4- ildehyde	3374 3374 2877	1660	1596	1381	1157 1099 1032	1565	1503 1475		ı	I.	ı	29-87°C 208-404°C 404-694°C	[4]
chitos quin	ian with 1-Phenyl-1H- oxaline-3-carbaldehyde	3374 3374 2877	1660	1596	1381	1157 1099 1032	1568	1505 1476	i.	I	ı	ı	28-98°C 207-434°C 434-695°C	[4]
chite 1H-	ssan with -1-phenyl-3- pyrazole-4-carbaldehy de	3439 3439 2900- e 2800	1644	I	1426	1057	1634	1400- 1500	1.23	1.91	8.78	8.38 7.87 7.70–7.08	~92°C ~294°C ~530°C	[5]
chit I-py	osan with 1- phenyl-3- razole -4-carbaldehyde	3439 3439 2900- 2800	1636	ı	1426	1157	1634	1400- 1500	1.23	1.91	9.72	10.17, 9.41 8.0, 7.5	~35°C ~295°C ~530°C	[5]
chit IH-J	osan with 1-phenyl-3- yyrazole-4-carbaldehyde	3439 3439 2900- 2800	1644	1	1426	1057	1634	1400- 1500	1.24	1.91	9.11	9.99, 9.39 8.67, 8.33 8.0, 7.5	~70°C ~304°C ~530°C	[5]

Table 1. Spectral and thermal data of Schiff bases of chitosan

	Ref		[9]	[9]	[9]	Ε	[7]	8	8	8	8	8	[6]
	TGA analysis	Temp. (°C	I			55-100°C 260-428°C t>430°C	ı	55-230°C 230-320°C 320-600°C	55-230°C 230-320°C 320-630°C	55-230°C 230-325°C 325-625°C	55-220°C 220-330°C 330-625°C	55-225°C 225-325°C 325-625°C	,
on)	(mqq/i	$\mathrm{H}^{-\mathrm{Ar}}$	7.3-7.6	7-7.3	6.6-7			7.2-7.3 6.8	7.2 6.8	7.4 7.2 6.9	9	7.3 7.1 6.9	6.8-7.6
	eaks (č	H- _{imine}	9.63	9.23	9.26	ı	I.	9.47	9.45	9.53	9.79	9.48	9.78
uati	MR p	Н-2	2.88	2.45	2.78	ı	i.	2.97	2.97	2.98	2.95	2.97	3.1
ntin	N H _I	H- _{amide}	1.77	1.34	1.68	,		1.87	1.87	1.86	1.85	1.87	·
<u></u> Ú	-	VC=C	1475	1475	1507	1466	I.	1520 1458	1520	1512 1458	1520 1458	1512 1458	1498
san	(₁	VC=N	1647	1598	1646	1646	1646	1636	1640	1641	1610	1640	1631
hitc	s (cm ⁻	Vglyc	1111	1057	1022	1160	1160	1026	1026	1026	1026	1026	,
ases of c	Main IR absorption band	VC-N	ı.	'	,	,		1	1	i.	i.	i.	'
		NNH	ı.	'	,	1600	1600	1589	1589	1589	1589	1589	1598
iff b		VC=0			ı	1690	1680	i.	,	ı.	ı.	ı.	1655
Sch		VC-H	2916	2874	2870	2984	2991			ı.	ı.	i.	2880
a of		VOH	3416	3423	3425	3400	3400	i.	i.	I.	I.	1	3368
l dat		VNH2	ī	'	·	3400	3400	ı	I	I	ı	I	3368
Table 1. Spectral and therm	Schiff base of chitosan		Schiff base of chitosan with benzaldehyde	Schiff base of chitosan with 4-(dimethylamino)benzaldehyde	Schiff base of chitosan with 4-hydroxy-3-methoxybenzaldehyde	Schiff base of chitosan with 5-Chloro-3-methyl-1-phenyl-1H- pyrazole-4- carbaldehyde	Schiff base of chitosan with 1-phenyl-3-methyl-5-p-nitro phenyl- 1H-pyrazole-4-carbaldehyde	Schiff base of chitosan with vanillin	Schiff base of chitosan with ethylvanilin	Schiff base of chitosan with veratraldehyde	Schiff base of chitosan with 2,4,6-trimethoxy benzaldehyde	Schiff base of chitosan with 4-ethoxy 3-hydroxy benzaldehyde	Schiff base of chitosan with salicylaldehyde
	No		6	10	11	12	13	14	15	16	17	18	19

2.Schiff bases of chitosan with antimicrobial activity

The antimicrobial activities of chitosan and its Schiff bases were tested against *Staphylococcus aureus*, *Bacillus cereus*, *Bacillus subtilis* as Gram-positive bacteria and *Escherichia coli*, *Pseudomonas aeruginosa*, *Salmonella sp*, *Klebsiella pneumonia as* Gram-negative bacteria. Antifungal activity was tested on *Candida albicans* and *Aspergillus niger*. The antimicrobial activities of the chitosan derivatives were compared with those of chitosan. The antimicrobial activities of the Schiff bases of chitosan were stronger than those of chitosan in most microorganisms.

The Schiff bases of chitosan with vanillin, ethyl vanillin, veratraldehyde, 2,4,6 trimethoxy benzaldehyde and 4-ethoxy-3-hydroxy showed antimicrobial benzaldehyde better activities Bacillus subtilis. Escherichia against *Staphylococcus* aureus, coli and Aspergillus niger than chitosan. Among all the above-mentioned chitosan derivatives, the Schiff base with vanillin was most effective against Staphylococcus aureus and Bacillus subtilis. Also, the Schiff bases of chitosan with vanillin and 4-ethoxy-3-hydroxy benzaldehyde showed the best antifungal activity against Aspergillus niger [8].

The chitosan Schiff bases prepared via coupling of chitosan with 4-chloro benzaldehyde and benzophenone [1], respectively, were more effective against Gram-negative bacteria than Gram-positive. Chitosan-4-chloro benzaldehyde Schiff base showed the highest activity against *Candida albicans*, *Pseudomonas aeruginosa* and *Bacillus cereus*. The activity of chitosan-benzophenone Schiff base against *Staphylococcus aureus*, *Bacillus cereus*, *Escherichia coli* and *Salmonella sp* was stronger than those of chitosan.

From the study of antimicrobial activity of the chitosan Schiff bases obtained by coupling of chitosan with indole-3-carboxaldehyde and 4-dimethylamino benzaldehyde, respectively, was observed the highest activity for the Schiff base of chitosan with indole-3carboxaldehyde [3]. However, the chitosan Schiff base with 4dimethylaminobenzaldehyde was more effective against *Staphylococcus aureus* and *Bacillus cereus* Gram-positive bacteria.

The inhibitory effect of chitosan Shiff bases resulted from the reaction of chitosan with 5-chloro-3-methyl-1-phenyl-1H-pyrazole-4-

1-phenyl-3-methyl-5-o-cresyloxy-1Hpyrazole-4carbaldehyde, carbaldehyde, 1-phenyl-3-methyl-5(-2-chloro phenyl)-1H-pyrazole-4carbaldehyde, 1-phenyl-3-methyl-5(-p-Nitro phenyl)-1H-pyrazole-4carbaldehyde and 3-methyl-1-phenyl-5-(piperidin-1-yl)-1H-pyrazole-4respectively, against Staphylococcus carbaldehyde, aureus, Bacillus subtilis and Escherichia coli was higher than those of chitosan. From the above-mentioned derivatives, 5-chloro-3- methyl-1-phenyl-1H-pyrazole-4-carbaldehyde showed better antibacterial and antifungal activity (against *Staphylococcus* Bacillus subtilis, Escherichia aureus, coli and Candida albicans) [7].

3. Pollutants removal with Schiff bases of chitosan

The Schiff bases of chitosan with 1*H*-pyrazolo[3,4-b]quinoxaline-3-carbaldehyde and 1-phenyl-1H-pyrazolo[3,4-b]quinoxaline-3carbaldehyde, respectively were evaluated for the removal of hexavalent chromium from synthetic samples [4]. The optimum pH value for the absorption process of Cr(VI) was 6. The two Schiff bases of chitosan showed high removal efficiency. The best removal efficiency for 1*H*-pyrazolo[3,4-b]quinoxaline-3-carbaldehyde was 96.78 % at the optimum dose of 150 mg/L. In case of 1-phenyl-1H-pyrazolo[3,4b]quinoxaline-3-carbaldehyde, the best removal efficiency was 98.85 % at the dose of 250 mg/L. From the study of the effect of chromium initial concentration it was observed that the removal efficiency decreased when the initial concentration of Cr(VI) increased.

The salicylaldehyde-chitosan Schiff base was used as an electropositive adsorbent in order to study the electrostatic adsorption behavior of dyes like methylorange, anionic xylenol orange, methylene blue and cationic basic fuchsin, respectively [9]. The results showed that the anionic dyes exhibited a much better adsorption performance than cationic dyes. The color of methylorange and xylenol orange (the anionic dyes) disappeared after 40 min while basic fuchsin and methylene blue (the cationic dyes) exhibited an insignificant adsorption performance.

4. CONCLUSION

Some chitosan's Schiff bases obtained by the reaction of C₂-NH₂ group with primary amines were characterized using FTIR and ¹H NMR spectroscopy and thermal analysis.

The antimicrobial activities of chitosan and its Schiff bases were studied against Staphylococcus aureus, Bacillus cereus, Bacillus subtilis, coli, Pseudomonas aeruginosa, Salmonella Escherichia sp, Klebsiella tested pneumonia. Antifungal activity was on Candida albicans and Aspergillus niger.

The Schiff bases of chitosan with 1*H*-pyrazolo[3,4-b]quinoxaline-3-carbaldehyde and 1-phenyl-1H-pyrazolo[3,4-b]quinoxaline-3carbaldehyde were studied for the removal of hexavalent chromium from synthetic samples and it was observed the best removal efficiency for 1*H*-pyrazolo[3,4-b]quinoxaline-3-carbaldehyde.

The Schiff base of chitosan with salicylaldehyde was used as an adsorbent in order to study the electrostatic adsorption behaviour of some dyes.

REFERENCES

- [1] T. M. Tamer, M. A. Hassan, A. M. Omer, W. M.A. Baset, M. E. Hassan, M. E.A. El-Shafeey, M. S. Mohy Eldin, *Process Biochemistry*, 51(10) (2016) 1721, https://doi.org/10.1016/j.procbio.2016.08.002
- [2] W. Wang, Q. Meng, Q. Li, J. Liu, M. Zhou, Z. Jin, K. Zhao, International Journal of Molecular Sciences, 21 (2020) 487, doi:10.3390/ijms21020487
- [3] M. A. Hassan, A. M. Omer, E. Abbas, W. M. A. Baset, T. M. Tame, Scientific Reports, 8 (2018) 11416, DOI:10.1038/s41598-018-29650-w 1
- M. Elhag, H. E. Abdelwahaba, M. A. Mostafa, A. Z. Nasr, M. M. El Sadek, *International Journal of Biological Macromolecules*, 163 (2020) 2180, https://doi.org/10.1016/j.ijbiomac.2020.09.090
- [5] A. A. Hamed, I. A. Abdelhamid, G. R. Saad, N. A. Elkady, M. Z. Elsabee, International Journal of Biological Macromolecules, 153 (2020) 492, https://doi.org/10.1016/j.ijbiomac.2020.02.302
- [6] J. Haque, V. Srivastava, D. S. Chauhan, H. Lgaz, M. A. Quraishi, ACS Omega, 3 (2018) 5654, DOI: 10.1021/acsomega.8b00455
- [7] S. M. Anush, B. Vishalakshi, B. Kalluraya, N. Manju, International Journal of Biological Macromolecules, <u>119</u> (2018) 446, https://doi.org/10.1016/j.ijbiomac.2018.07.129
- [8] S. Lal, S. Arora, C. Sharma, Journal of Thermal Analysis and Calorimetry, 124 (2016) 909, https://doi.org/10.1007/s10973-015-5227-3
- [9] C. Huang, H. Liao, X. Ma, M. Xiao, X. Liu, S. Gong, X. Shu, X. Zhou, Chemical Physics Letters, 780 (2021) 138958, <u>https://doi.org/10.1016/j.cplett.2021.138958</u>